

## Constituents from the Leaves of *Aristolochia elegans*

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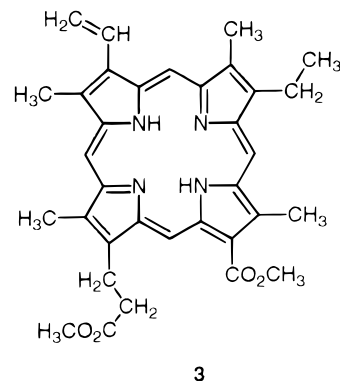
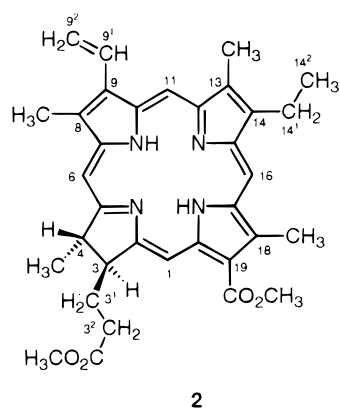
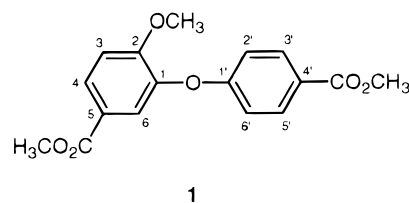
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One new biphenyl ether, aristogin C (**1**), and two new porphyrins, aristophylls A (**2**) and B (**3**), as well as 11 known compounds, were isolated from the leaves of *Aristolochia elegans*. Their structures were elucidated according to the spectroscopic (NMR and MS) analyses or by comparison with literature values.

*Aristolochia elegans* Mast. (Aristolochiaceae) is a perennial shrub cultivated as an ornamental plant in Taiwan.<sup>1</sup> Several reports have been found on the isolation of lignans, diterpenoids, and alkaloids from the leaves, stems, and roots of this plant.<sup>2–8</sup> In a continuing search for novel bioactive compounds from the genus *Aristolochia*,<sup>9</sup> the leaves of *A. elegans* were investigated. Three new compounds—one biphenyl ether, aristogin C (**1**), and two porphyrins, aristophylls A (**2**) and B (**3**)—and 11 known compounds were isolated from the hot methanolic extract. Here we report the structure elucidation of **1–3** by spectroscopic (NMR and MS) analyses.

Aristogin C (**1**), C<sub>17</sub>H<sub>16</sub>O<sub>6</sub>, was obtained as a colorless oil. There were two sets of mutually coupled protons in the aromatic region of the <sup>1</sup>H NMR spectrum of **1**: one at δ 7.04 (d, *J* = 8.7 Hz, H-3), 7.74 (d, *J* = 1.7 Hz, H-6), and 7.93 (dd, *J* = 8.7, 1.7 Hz, H-4) indicated a 1,2,5-trisubstituted benzene; the other at δ 6.90 (2H, d, *J* = 8.8 Hz, H-2' and H-6') and 7.98 (2H, d, *J* = 8.8 Hz, H-3' and H-5') indicated a 1',4'-disubstituted benzene. A strong carbonyl band in the IR spectrum and two ester carbonyl signals at δ 166.1 and 166.6 in the <sup>13</sup>C NMR spectrum indicated that two of the three methoxyl singlets were carbomethoxy groups. Consequently, the two benzene rings are linked by an oxygen, typical of a diphenyl ether. The regiochemistry of the substituents, two carbomethoxyls and one methoxyl, was confirmed by NOEs between δ 7.04 (H-3) and 3.88 (OMe) and 7.93 (H-4) in a 2D NOESY experiment, which led us to place a carbomethoxyl group on C-5 and to assign structure **1** to aristogin C.

Aristophyll A (**2**) was isolated as violet granules that exhibited NMR data similar to those of methyl pheophorbide-a.<sup>10</sup> UV absorption bands at 221, 282, 305, 369 (sh), 398, 498, 527, 607, and 664 nm suggested that **2** was a chlorophyll derivative.<sup>11</sup> The FABMS quasi molecular ion at *m/z* 567 [M + H]<sup>+</sup> indicated the molecular formula C<sub>34</sub>H<sub>38</sub>N<sub>4</sub>O<sub>4</sub>. Broad IR absorption at 3327 cm<sup>-1</sup> and the broad <sup>1</sup>H NMR signal at δ 1.64 (2H, D<sub>2</sub>O exchangeable) showed the presence of two typical upfield NH protons in the porphyrin ring. The IR spectrum also showed carbonyl absorptions at 1740 and 1699 cm<sup>-1</sup>. As in the aliphatic region of the <sup>1</sup>H NMR spectrum of **2**, a doublet methyl at δ 1.88 (*J* = 7.2 Hz, 4-Me) coupled with a downfield methine proton at δ 4.51 (m, H-4) were typical of a partially reduced dihydroporphyrin. In addition, an ethylene group at δ 2.38 and 2.55 (m, each 1H, H-3<sup>2</sup>), as well as 2.47 and 2.59 (m, each 1H, H-3<sup>1</sup>), along with a carbomethoxyl, constructed a



methyl propanoate side chain attached to C-3. The characteristic ethyl group at δ 1.72 and 3.78, together with a vinyl substituent at δ 6.14, 6.33, and 8.07, were placed on C-14 and C-9, respectively. Three methyl singlets at δ 3.30 (13-Me), 3.46 (8-Me), and 3.79 (18-Me) and four aromatic singlets at δ 8.73 (H-6), 9.61 (H-11), 9.75 (H-16), and 9.80 (H-1) represent the protons outside the dihydroporphyrin ring that caused these downfield shifted signals. Therefore, the last carbomethoxyl group was placed on C-19. All of the <sup>1</sup>H and <sup>13</sup>C NMR assignments were confirmed by COSY, HMQC, NOESY, and HMBC experiments. Thus, structure **2** was assigned to aristophyll A.

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Aristophyll B (**3**),  $C_{34}H_{36}N_4O_4$ , had two fewer hydrogens than **2**. The spectral data of **3** were almost the same as those of **2**. Four downfield methyl groups in **3**, instead of three, suggested that it consisted of a porphyrin ring. The NOE experiment confirmed the locations of substituents as shown. Consequently, aristophyll B was established as **3**, a compound previously synthesized by Conant and Bailey.<sup>12</sup>

The known compounds phytol,<sup>13</sup>  $\beta$ -sitosterol,<sup>13</sup> (-)-pinoresinol,<sup>14</sup> methylparaben,<sup>15</sup> *p*-methylvanillate,<sup>16</sup> *p*-hydroxybenzoic acid,<sup>15</sup> corydaldine,<sup>17</sup> thalifoline,<sup>18</sup> northalifoline,<sup>19</sup> *N*-methylcorydaldine,<sup>20</sup> and *N*-methyl-6,7-dimethylisoquinoline<sup>21</sup> were also isolated and characterized by comparison of their spectroscopic data (UV, IR, NMR, and MS) with literature values. In this study, we have isolated five 1-quinolones and a biphenyl ether, which may represent metabolites of bisbenzylisoquinoline alkaloids in the plant.<sup>22</sup>

## Experimental Section

**General Experimental Methods.** Melting points were not corrected. UV spectra were recorded in MeOH. IR spectra were recorded as KBr disks. NMR spectra were recorded at 200 or 400 MHz for <sup>1</sup>H and 50 or 100 MHz for <sup>13</sup>C; all chemical shifts are reported in parts per million ( $\delta$ ) from TMS as an internal standard. Mass spectra were performed in the EI or FAB (matrix: glycerol) mode.

**Plant Material.** *A. elegans* Mast. was collected in May 1992, from Tainan Hsien, Taiwan, and verified by Prof. C. S. Kuoh. A specimen of the plant (NCKU Wu 92008) has been deposited at the herbarium of National Cheng Kung University, Tainan, Taiwan.

**Extraction and Separation.** Fresh leaves of *A. elegans* (1.1 kg) were extracted with hot MeOH ( $\times 5$ ) and concentrated to give a dark brown syrup that was partitioned between H<sub>2</sub>O and CHCl<sub>3</sub>, and then *n*-butanol. This resulted in CHCl<sub>3</sub>, *n*-butanol, H<sub>2</sub>O, and insoluble portions after evaporation of the solvent. The CHCl<sub>3</sub> portion was chromatographed over Si gel using a gradient of *n*-hexane and Me<sub>2</sub>CO to afford 11 fractions. Fraction 7 was chromatographed over silica gel using CHCl<sub>3</sub>-Me<sub>2</sub>CO and rechromatographed by preparative TLC to yield phytol (68.2 mg), **1** (3.4 mg), and  $\beta$ -sitosterol (648.2 mg). Using the same procedure, fraction 8 yielded methylparaben (1.8 mg), *p*-methylvanillate (1.0 mg), *p*-hydroxybenzoic acid (1.0 mg), **2** (6.2 mg), and **3** (5.3 mg). Fraction 10 gave corydaldine (2.3 mg), *N*-methyl-6,7-dimethylisoquinoline (3.7 mg), (-)-pinoresinol (1.6 mg), and thalifoline (2.1 mg). The combined *n*-butanol and insoluble portions were chromatographed on a cation exchange column eluted with 5% NH<sub>4</sub>OH to give an alkaloid fraction. Repeated chromatography on a C<sub>18</sub> column produced thalifoline (2.2 mg), *N*-methylcorydaldine (1.2 mg), and northalifoline (1.9 mg).

**Aristogin C (1):** colorless oil; UV  $\lambda_{\max}$  (log  $\epsilon$ ) 255 (4.82), 286 (4.09, sh) nm; IR  $\nu_{\max}$  1645, 1610, 1510 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  3.85 (3H, s, OMe), 3.88 (3H, s, OMe), 3.89 (3H, s, OMe), 6.90 (2H, d,  $J$  = 8.8 Hz, H-2' and H-6'), 7.04 (1H, d,  $J$  = 8.7 Hz, H-3), 7.74 (1H, d,  $J$  = 1.7 Hz, H-6), 7.93 (1H, dd,  $J$  = 8.7, 1.7 Hz, H-4), 7.98 (2H, d,  $J$  = 8.8 Hz, H-3' and H-5'); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  52.0, 52.1, 56.1, 112.1, 115.9, 123.3, 123.4, 124.3, 128.1, 131.6, 143.1, 155.5, 161.7, 166.1, 166.6; EIMS  $m/z$  316 (M<sup>+</sup>, 100), 285 (85), 183 (10), 127 (24), 104 (7), 77 (12); HREIMS  $m/z$  316.0946 (calcd for C<sub>17</sub>H<sub>16</sub>O<sub>6</sub>, 316.0948).

**Aristophyll A (2):** violet granules (Et<sub>2</sub>O), mp 261–262 °C; [ $\alpha$ ]<sub>D</sub> -238.3° (c 0.06, CHCl<sub>3</sub>); UV  $\lambda_{\max}$  221, 282, 305, 369 (sh), 398, 498, 527, 607, 664 nm; IR  $\nu_{\max}$  3327, 1740, 1699, 1616, 1506 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  1.64 (2H, br s, D<sub>2</sub>O exchangeable, 2  $\times$  NH), 1.72 (3H, t,  $J$  = 7.2 Hz, H-14<sup>2</sup>), 1.88 (3H, d,  $J$

= 7.2 Hz, 4-Me), 2.38 and 2.55 (each 1H, m, H-3<sup>2</sup>), 2.47 and 2.59 (each 1H, m, H-3<sup>1</sup>), 3.30 (3H, s, 13-Me), 3.46 (3H, s, 8-Me), 3.61 (3H, s, OMe), 3.78 (2H, q,  $J$  = 7.2 Hz, H-14<sup>1</sup>), 3.79 (3H, s, 18-Me), 4.35 (3H, s, OMe), 4.48 (1H, m, H-3), 4.51 (1H, m, H-4), 6.14 (1H, d,  $J$  = 10.0 Hz, H-9<sup>2</sup>), 6.33 (1H, d,  $J$  = 16.4 Hz, H-9<sup>2</sup>), 8.07 (1H, dd,  $J$  = 16.4, 10.0 Hz, H-9<sup>1</sup>), 8.73 (1H, s, H-6), 9.61 (1H, s, H-11), 9.75 (1H, s, H-16), 9.80 (1H, s, H-1); <sup>13</sup>C NMR (CDCl<sub>3</sub>)  $\delta$  11.4 (3-Me), 12.1 (8-Me), 13.7 (8-Me), 17.6 (C-14<sup>2</sup>), 19.7 (C-14<sup>1</sup>), 23.4 (4-Me), 31.1 (C-3<sup>2</sup>), 32.4 (C-3<sup>1</sup>), 48.6 (C-4), 51.6 (OMe), 51.9 (OMe), 54.7 (C-3), 93.0 (C-6), 96.3 (C-1), 98.9 (C-11), 102.1 (C-16), 119.0 (C-19), 121.9 (C-9<sup>2</sup>), 129.4 (C-9<sup>1</sup>), 130.2 (C-8), 130.5 (C-17), 134.7 (C-9), 135.1 (C-10), 136.1 (C-13), 137.2 (C-2), 140.0 (C-18), 140.3 (C-7), 144.9 (C-14), 149.4 (C-15), 154.1 (C-12), 166.7 (C-20), 167.1 (C=O), 171.3 (C-5), 173.9 (C=O); FABMS  $m/z$  567 [M + H]<sup>+</sup> for C<sub>34</sub>H<sub>38</sub>N<sub>4</sub>O<sub>4</sub> (100), 537 (11), 493 (10), 307 (12), 176 (28), 154 (83), 136 (71), 107 (32).

**Aristophyll B (3):** violet granules (Et<sub>2</sub>O), mp 269–271 °C; UV  $\lambda_{\max}$  264, 389, 404, 512, 553, 633 nm; IR  $\nu_{\max}$  3300, 1715, 1695 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>)  $\delta$  4.31 (2H, br s, D<sub>2</sub>O exchangeable, 2  $\times$  NH), 1.81 (3H, t,  $J$  = 7.6 Hz, H-14<sup>2</sup>), 3.26 (2H, t,  $J$  = 7.6 Hz, H-3<sup>2</sup>), 3.51 (3H, s, 4-Me), 3.52 (3H, s, 13-Me), 3.53 (3H, s, 8-Me), 3.68 (3H, s, OMe), 3.86 (3H, s, 18-Me), 3.97 (2H, q,  $J$  = 7.6 Hz, H-14<sup>1</sup>), 4.33 (2H, t,  $J$  = 7.6 Hz, H-3<sup>1</sup>), 4.44 (3H, s, OMe), 6.13 (1H, d,  $J$  = 11.2 Hz, H-9<sup>2</sup>), 6.26 (1H, d,  $J$  = 16.0 Hz, H-9<sup>2</sup>), 8.11 (1H, dd,  $J$  = 16.0, 11.2 Hz, H-9<sup>1</sup>), 9.69 (1H, s, H-6), 9.85 (1H, s, H-11), 9.89 (1H, s, H-16), 10.82 (1H, s, H-1); FABMS  $m/z$  565 [M + H]<sup>+</sup> for C<sub>34</sub>H<sub>38</sub>N<sub>4</sub>O<sub>4</sub> (67), 491 (6), 307 (11), 176 (15), 154 (100), 136 (78), 107 (37).

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